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Biological Barriers Research Group

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RESEARCH AREA

Organisms are protected by biological barriers from harmful effects. These barriers also impede drug penetration. Our lab investigates methods to increase drug delivery on culture models of the blood-brain, nasal, corneal, respiratory and intestinal barriers. The pathways examined are (i) reversible opening of tight intercellular junctions by peptides or small molecules; (ii) targeting solute carriers at barriers for drug delivery by nanoparticles. Cellular toxicity of active ingredients and pharmacautical excipients are measured by a real-time impedance-based method. Double and triple co-culture models are used for experiments, and a microfluidic integrated chip has been developed in a collaborative project. Our other major research interest is the examination of blood-brain barrier injury and dysfunctions in different diseases, like Alzheimer's disease, acute pancreatitis and diabetes. The goal of these experiments is to reveal the effect of disease pathogenic factors on blood-brain barrier functions and to identify protective molecules. The protection of brain endothelial cells and the improvement of BBB functions in pathological conditions, the exploration of new approaches for drug transport/targeting to brain may have therapeutic potential in the treatment of central nervous system diseases.

TECHNIQUES AVAILABLE IN THE LAB

Mammalian cell culture; primary cultures from brain and brain microvessels; models of biological barriers by double and triple co-cultures; cell culture models in microfluidic chips; electric resistance measurements of cell layers; permeability of drugs across culture models; immunohistochemistry; phase contrast, fluorescent and confocal microscopy; ELISA; measurement of nitric oxide and reactive oxygen species production in cells; colorimetric and impedance-based toxicity tests.

SELECTED PUBLICATIONS

Mészáros, M., Porkoláb, G., Kiss, L., Pilbat, A.M., Kóta, Z., Kupihár, Z., Kéri, A., Galbács, G., Siklós, L., Tóth, A., Fülöp, L., Csete, M., Sipos, Á., Hülper, P., Sipos, P., Páli, T., Rákhely, G., Szabó-Révész, P., **Deli, M.A.**, Veszelka, S. (2018) Niosomes decorated with dual ligands targeting brain endothelial transporters increase cargo penetration across the blood-brain barrier. **Eur J Pharm Sci 123:** 228-240.

Veszelka, S., Tóth, A., Walter, F.R., Tóth, A.E., Gróf, I., Mészáros, M., Bocsik, A., Hellinger, É., Vastag, M., Rákhely, G., **Deli, M.A.** (2018) Comparison of a rat primary cell-based blood-brain barrier model with epithelial and brain endothelial cell lines: gene expression and drug transport. **Front Mol Neurosci 11:** 166.

Walter, F.R., Valkai, S., Kincses, A., Petneházi, A., Czeller, T., Veszelka, S., Ormos, P., **Deli, M.A.,** Dér, A. (2016) Versatile lab-on-a-chip tool for modeling biological barriers. **Sens Actuators B Chem 222:** 1209-1219.

Bocsik, A., Walter, F.R., Gyebrovszki, A., Fülöp, L., Blasig, I., Dabrowski, S., Ötvös, F., Tóth, A., Rákhely, G., Veszelka, S., Vastag, M., Szabó-Révész, P., **Deli, M.A.** (2016) Reversible opening of intercellular junctions of intestinal epithelial and brain endothelial cells with tight junction modulator peptides. **J Pharm Sci 105:** 754-765.

Veszelka, S., Tóth, A.E., Walter, F.R., Datki, Z., Mózes, E., Fülöp, L., Bozsó, Z., Hellinger, E., Vastag, M., Orsolits, B., Környei, Z., Penke, B., **Deli, M.A.** (2013) Docosahexaenoic acid reduces amyloid-β induced toxicity in cells of the neurovascular unit. **J Alzheimers Dis 36:** 487-501.